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Global prediction of planktic foraminiferal fluxes from hydrographic and productivity data

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Abstract. Understanding and quantifying the seasonal and spatial distribution of planktic foraminiferal fluxes reflected in sedimentary assemblages is key to interpret foraminifera-based proxies in paleoceanography. Towards this goal we present an empirical model to predict foraminiferal fluxes on a global scale.

A compilation of planktic foraminiferal flux and export production data from globally distributed sediment traps together with environmental data of sea-surface temperature and mixed-layer depth from online databases is used to calibrate the model that calculates monthly foraminiferal fluxes for the 18 most common species. The calibrated model is then forced with a global data set of hydrographic and productivity data to predict monthly foraminiferal fluxes worldwide. The predictive skills of the model are assessed by comparing the model output with planktic foraminiferal assemblages from globally distributed surface sediments as well as with measured foraminiferal fluxes of sediment traps not included in the calibration data set.

Many general distribution patterns of foraminiferal species recognized from the model output compare favorably with observations from coretops or sediment traps, even though the model still produces problematic results in some places. Among others, meridional gradients in species richness and diversity, increased relative abundances of *Neogloboquadrina pachyderma* (dex.) in upwelling areas, and peak abundances of *Globigerinella siphonifera* in oligotrophic subtropical gyres show good agreement between model and coretops. Absolute foraminiferal fluxes are significantly underestimated in most cases, while seasonal variations can be reproduced for some species. Interannual differences in foraminiferal fluxes are not reflected by the model which might partly be due to a lack of actual environmental data for the calibration and model experiments.

The limited predictive skills of the model suggest that additional parameters should be considered. Export production should probably be replaced by a more realistic representation of food availability for planktic foraminifera. This could be achieved by adding a dynamic component to the model and linking it to an ecosystem model.

1 Introduction

The shell chemistry as well as the assemblage composition of planktic foraminifera are among the most important proxies used in paleoceanography (e.g. Wefer et al., 1999). However, there is often a significant seasonal bias in foraminiferal species fluxes to the seafloor, which is crucial for paleoceanographic interpretations of the proxy data (e.g. Deuser and Ross, 1989; Wefer, 1989; Mulitza et al., 1998; Ganssen and Kroon, 2000; King and Howard, 2001; Pflaumann et al., 2003; Waelbroeck et al., 2005). For example, Niebler et al. (2003) suggested, that discrepancies in temperature reconstructions applying foraminifera- versus alkenone-based proxies might be due to different ecological and thus seasonal preferences of the shell producers.

Seasonal variations in foraminiferal fluxes are strongly influenced by the different species sensitivities to environmental parameters: Numerous studies on plankton-tow and sediment-trap material as well as on laboratory cultures and surface sediments have shown, that sea-surface temperature (SST), the thermal structure of the water column and food supply among others can have a considerable effect on fluxes and relative abundances of foraminiferal species (e.g. Bijma et al., 1990b; Ortiz et al., 1995; Watkins et al., 1996; Watkins and Mix, 1998; Eguchi et al., 1999; Schiebel et al., 2001; King and Howard, 2003a; Morey et al., 2005; Žarić et al., 2005, and references in Table 1).

Sediment traps are a useful tool in these investigations, as they can resolve seasonal and interannual differences in

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particle flux (e.g. Deuser, 1986; Fischer and Wefer, 1996; Kincaid et al., 2000). Because of the relatively large size and weight of planktic foraminifera, their shells usually reach the traps within days and mostly without significant lateral displacement (e.g. Takahashi and Bé, 1984). Hence, trap data can be directly related to modern surface hydrography (e.g. Tedesco and Thunell, 2003; Marchant et al., 2004; Mohiuddin et al., 2004; Žarić et al., 2005).

To minimize errors in the application of foraminifera-based proxies it is of great importance to thoroughly understand the seasonal and spatial distribution of foraminiferal fluxes reflected in sedimentary assemblages and to be able to quantify it on a global scale.

Here, we present, to our knowledge, the first attempt to globally predict foraminiferal fluxes at species level (18 most common species) depending on the environmental parameters sea-surface temperature, mixed-layer depth (MLD) and export production in 1000 m water depth (PEX). The empirical model is calibrated by a compilation of planktic foraminiferal flux and PEX data from globally distributed sediment traps in combination with environmental data of SST and MLD from online databases. We force the calibrated model with a global data set of hydrographic and productivity data to calculate monthly foraminiferal fluxes on a global scale. We then compare modeled annual relative species abundances with core-top-derived foraminiferal assemblage data. Finally, we test the model by comparing predicted and measured foraminiferal fluxes at trap positions not included in the calibration.

Our study shows that the empirical model is able to reproduce many general distribution patterns of planktic foraminiferal assemblages observed in the world's oceans like diversity patterns or relative abundance distributions of some species, even though it cannot reliably predict absolute monthly foraminiferal fluxes yet.

2 Material and methods

2.1 Calibration data set

To calibrate our model we used a global data set compiled by Žarić et al. (2005). This database contains planktic foraminiferal fluxes calculated from various sediment-trap investigations, actual SSTs taken from the IGOSS database (Reynolds and Smith, 1994) (<http://iridl.ldeo.columbia.edu/SOURCES/IGOSS/.nmc/>) as well as export production data, which have been derived from organic carbon fluxes measured with the same traps. To account for remineralization during descent, the organic carbon fluxes have been normalized to an average trap depth of 1000 m by applying the power function proposed by Martin et al. (1987) with correction after Francois et al. (2002; Eq. 14 therein). Even though this can only be a first approximation (Žarić et al., 2005), we decided to still use export production, because it

was the only productivity-related parameter being available in the temporal resolution of sediment trap samples.

We extended the database of Žarić et al. (2005) by adding several North Atlantic traps (Jensen, 1998; Peinert et al., 2000; Schröder-Ritzrau et al., 2000; Antia et al., 2001; Schiebel, 2002) complementing SST and PEX data for every sample as described by Žarić et al. (2005). Furthermore we added information on the depths of the mixed layer. These were calculated from monthly temperature and salinity data obtained from the World Ocean Atlas 2001 (WOA 2001; Conkright et al., 2002) (<http://www.nodc.noaa.gov/OC5/WOA01/>) based on a constant density difference criterion of 0.125 sigma units between ocean surface and base of the mixed layer. In polar regions, where deep mixing occurs, a density difference of 10^{-5} sigma units was used. The minimum mixed-layer depth was set to 25 m. The same value was used for sea-ice covered regions. Data from the $1 \times 1^\circ$ grid-points closest to each sediment trap position were transformed to weekly values, and mean mixed-layer depths were calculated for every sampling interval. We applied the same time-lag between trap data and surface hydrography as described by Žarić et al. (2005): Due to life cycles of foraminifera an adjustment of two weeks was used for all sediment-trap data (Sautter and Thunell, 1991). An additional one-week correction (i.e. 3 weeks in total) was used for traps deeper than 1750 m water depth to account for foraminiferal settling through the water column, which translates into assumed average sinking velocities of 500 m per day (Takahashi and Bé, 1984). Furthermore, where only fluxes of *all* occurring species were given, we set fluxes of the remaining species to zero ("void observations"). If no information was available on the organic carbon flux or if the total foraminiferal flux was zero, the sample was excluded from further investigation. Altogether the calibration data set consists of 1327 samples. Table 1 summarizes locations, details and references of the sediment trap studies included in our model calibration.

2.2 Statistical analysis and model development

Foraminiferal flux data of the following 18 species were examined in our study: *Globigerina bulloides*, *Globigerinella calida*, *G. siphonifera*, *Globigerinita glutinata*, *Globigerinoides ruber* (white and pink varieties), *G. sacculifer*, *Globorotalia inflata*, *G. menardii*, *G. scitula*, *G. truncatulinoides*, *Globoturborotalita rubescens*, *Neogloboquadrina dutertrei*, *N. pachyderma* (sinistral and dextral coiling varieties), *Orbulina universa*, *Pulleniatina obliquiloculata* and *Turborotalita quinqueloba*. Each of these species was present in at least 15% of the samples. Other species account for only 5.6% of the foraminiferal assemblage on an average.

The empirical model is based on a multiple linear regression approach. As there is no linear relationship between most foraminiferal fluxes and environmental parameters, we applied the ACE algorithm prior to regression to estimate

Table 1. Locations, trap and water depths, sieve size and data sources of the planktic foraminiferal faunas included in the calibration data set.

Trap Location		Latitude [° N]	Longitude [° E]	Trap Depths [m]	Water Depth [m]	Sieve Size [μm]	References
Ocean Station Papa		50.00	−145.00	3800	4240	> 125	Reynolds and Thunell (1985, 1986); Sautter and Thunell (1989); Wong et al. (1999)
California Current	NS	42.09	−125.77	1000	2829	> 150	Ortiz and Mix (1992); Lyle et al. (1992)
	MW	42.19	−127.58	1000	2830	> 150	
	G	41.54	−132.02	1000	3664	> 150	
Peru-Chile Current ^a		−30.01	−73.18	2318	4345	> 150	Marchant et al. (1998); Hebbeln et al. (2000)
N' North Atlantic	OG	72.38	−7.71	500; 1000; 2300	2624	> 125	Jensen (1998); Peinert et al. (2000)
	NB	69.69	0.48	500; 1000	3254	> 125	
North Atlantic	L2 ^a	47.78	−19.78	1015; 2015; 3515	5483	> 150	Schiebel (2002); Antia et al. (2001)
	L3	54.66	−21.23	2200	5370	> 150	
Sargasso Sea		32.08	−64.25	3200	4200	> 125	Deuser et al. (1981); Deuser (1987); Deuser and Ross (1989)
Canary Islands	LP	29.76	−17.95	900	4327	> 125	Freudenthal et al. (2001);
	CI	29.18	−15.45	500; 750	3610	> 125	Abrantes et al. (2002);
	EBC	28.71	−13.16	700	996	> 125	Wilke et al. (2005) ¹
Cape Blanc	1	20.76	−19.74	2195	3646	> 150	Fischer and Wefer (1996);
	2–5 ^a	21.15	−20.68	732; 3552	4103	> 150	Fischer et al. (1996); Žarić et al. (2005)
W Equatorial Atlantic	1	−4.00	−25.57	652; 1232; 4991	5530	> 150	Fischer and Wefer (1996),
	2–3 ^a	−7.52	−28.04	631; 5031	5570	> 150	Fischer (unpubl. data); Žarić et al. (2005)
W Atlantic	WAB ^a	−11.57	−28.53	719; 4515	5472	> 150	Fischer (unpubl. data); Žarić et al. (2005)
E Equatorial Atlantic	1	3.17	−11.25	984	4524	> 150	Fischer and Wefer (1996); Žarić et al. (2005)
	2	1.78	−11.25	953	4399	> 150	
	3	−0.08	−10.77	1097	4141	> 150	
	4	−2.19	−10.09	1068	3906	> 150	
Walvis Ridge	2–3	−20.05	9.16	599; 1648	2202	> 150	Fischer and Wefer (1996);
	4	−20.13	8.96	1717	2263	> 150	Žarić et al. (2005)
Weddell Sea	1	−62.44	−34.76	863	3880	> 125	Donner and Wefer (1994)
	2–4 ^a	−64.91	−2.55	356; 4456	5032	> 125	
Arabian Sea	WAST ^a	16.33	60.49	3026	4016	> 150	Curry et al. (1992);
	CAST ^a	14.49	64.76	2909	3901	> 150	Guptha and Mohan (1996);
	EAST ^a	15.48	68.74	2775	3774	> 150	Haake et al. (1993)
Bay of Bengal	NBBT	17.45	89.60	967; 1498; 2029	2263	> 150	Guptha and Mohan (1996);
	CBBT	13.15	84.35	950; 2286	3259	> 150	Guptha et al. (1997);
	SBBT ^b	5.00	87.05	1518	4017	> 125	Unger et al. (2003)
NW' Pacific	WCT-1 ^a	25.00	136.99	917; 1388; 4336; 4758	5107	> 125	Mohiuddin et al. (2002)
	WCT-2 ^a	39.01	147.00	1371; 1586; 4787	5339	> 125	
NW' North Pacific	50N	50.02	165.03	3260	5570	> 125	Kuroyanagi et al. (2002)
	KNOT	43.97	155.05	2957	5370	> 125	
	40N	40.00	165.00	2986	5483	> 125	
Subantarctic Zone	SAZ 47	−46.76	142.07	1060; 3850	4540	> 150	King and Howard (2003a,b);
	SAZ 51	−51.00	141.74	3080	3780	> 150	Trull et al. (2001)
	SAZ 54	−53.75	141.76	830; 1580	2280	> 150	
Chatham Rise	NCR	−42.70	178.63	300; 1000	1500	> 150	King and Howard (2001);
	SCR	−44.62	178.62	300; 1000	1500	> 150	Nodder and Northcote (2001)

^a Position and/or depths averaged over more than one collection period^b Flux data available for *G. bulloides* only

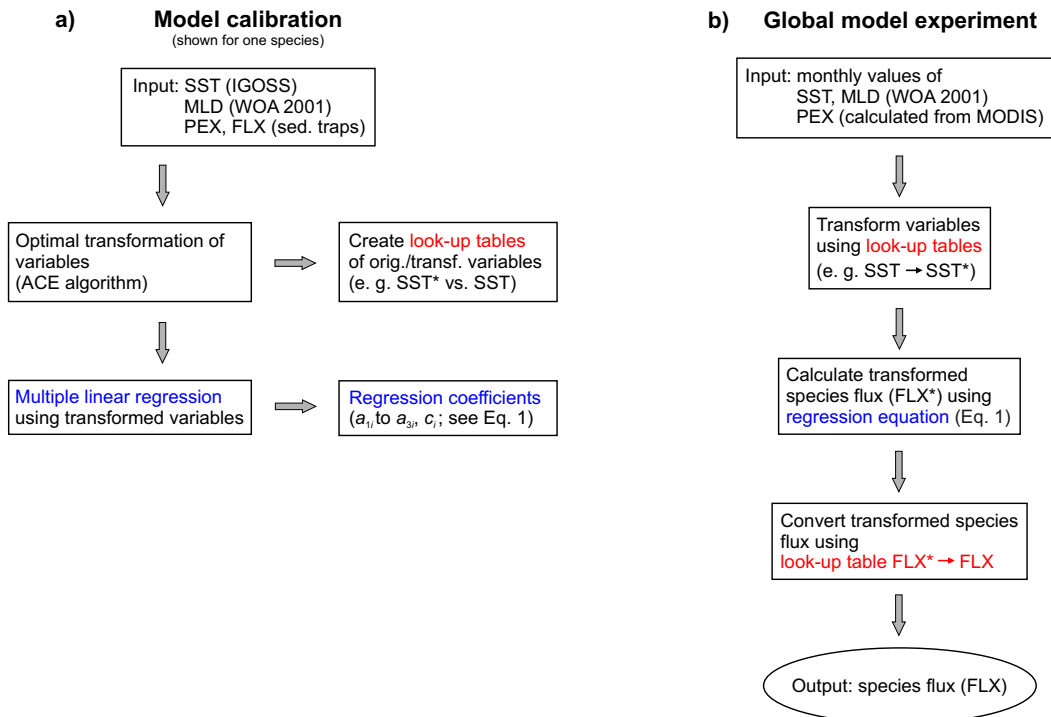


Fig. 1. Schematic summary of (a) the model calibration and (b) the global model experiment. SST = sea-surface temperature, MLD = mixed-layer depth, PEX = export production in 1000 m water depth, FLX = flux of a foraminiferal species. Note that actual SST data from the time of trap deployment were used for the calibration in (a), because these are more accurate when related to trap data. On the other hand, the global model experiment in (b) was forced with long-term monthly mean values of SST in order to better compare the model results to core-top samples. For further explanation, see text.

optimal transformations for multiple regression and correlation (Breiman and Friedman, 1985). The algorithm transforms each variable such, that multiple linear regression of the transformed variables (denoted by an asterisk) yields the highest coefficients of determination (r^2). For the statistical analysis being the basis of the model calibration we treated SST, MLD and PEX as predictor variables and the flux of the respective foraminiferal species (FLX) as the dependent variable:

$$\text{FLX}_i^* = a_{1i} \text{SST}^* + a_{2i} \text{MLD}^* + a_{3i} \text{PEX}^* + c_i. \quad (1)$$

Here SST^* , MLD^* and PEX^* are transformed values of sea-surface temperature, mixed-layer depth, and export production in 1000 m water depth, respectively. FLX_i^* is the transformed flux of species i ($i=1, \dots, 18$), a_{1i} to a_{3i} are species-dependent regression coefficients and c_i a regression constant. It should be mentioned, that since the environmental parameters can be partially intercorrelated, multicollinearity may have an effect on the statistical analysis.

Look-up tables were derived from the ACE output to be used for variable transformations within the model ($\text{SST} \rightarrow \text{SST}^*$, $\text{MLD} \rightarrow \text{MLD}^*$, $\text{PEX} \rightarrow \text{PEX}^*$) and regression parameters were estimated (see Fig. 1a). The calculated transformed species flux from Eq. (1) is back-transformed

($\text{FLX}^* \rightarrow \text{FLX}$) to obtain a flux in units of $[\text{ind. m}^{-2} \text{ d}^{-1}]$. The inverse transformation necessitates that the transformation of species fluxes with the ACE algorithm is forced to behave strongly monotonic, so that each transformed flux corresponds to a single untransformed value. Planktic foraminiferal species were analyzed separately on the dependence of their fluxes on the mentioned environmental parameters. For every analysis only those samples from the compilation were included, where all variables were available. Hence, the number of samples varied for every single species under consideration.

In total, the foraminiferal flux model comprises 54 look-up tables that allow for a transformation of the three environmental input parameters SST, MLD and PEX for each of the 18 species considered. Furthermore, it contains 18 look-up tables that permit back-transformation into single foraminiferal species fluxes. By forcing the model with a data set of the environmental input parameters, planktic foraminiferal species fluxes are calculated separately and these are additionally converted to percentages of the modeled foraminiferal assemblage. *G. ruber* (pink) is only calculated for the Atlantic Ocean, since it became extinct in the Indo-Pacific at 128 000 yr BP (Ivanova et al., 2003, see also Thompson et al., 1979). According to the SST ranges of the

Table 2. Locations, trap and water depths, sieve size and data sources of the planktic foraminiferal faunas not included in the calibration data set. These were used to compare the seasonal signal of measured and modeled foraminiferal fluxes.

Trap Location		Latitude [° N]	Longitude [° E]	Trap Depths [m]	Water Depth [m]	Sieve Size [μm]	References
San Pedro Basin		33.55	−118.50	500	880	>125	Sautter and Thunell (1991)
Greenland Sea		75.00	0.00	300; 900	3720	>125	Jensen (1998)
Cariaco Basin		10.50	−64.67	275	1400	>125	Tedesco and Thunell (2003)
Benguela Upwelling	WB1	−23.03	12.44	968	1803	>150	Žarić et al. (2005)
	Benguela	−23.00	12.98	545	595	>125	Giraudeau et al. (2000)
Arabian Sea	MST8-B	10.76	51.94	1265	1533	>150	Conan and Brummer (2000); Conan et al. (2002)
	WAST ^a	16.31	60.47	1028	4014	>150	Curry et al. (1992)
	CAST ^a	14.49	64.76	733	3901	>150	
	EAST ^a	15.48	68.74	1401	3776	>150	
	SAST ^b	13.13	67.12	1654	4075	>125	Guptha and Mohan (1996)
Subantarctic Zone	SAZ 47	−46.76	142.07	3850	4540	>150	King and Howard (2003a, b)

^a Position and depths averaged over more than one collection period^b Flux data available for *G. bulloides* only

individual species, which were derived as outlined in Žarić et al. (2005), cutoffs were defined to assure that species are not calculated out of their present-day SST range (Appendix A, Table A1).

2.3 Global model run

For a global model run the foraminiferal flux model was forced with a global monthly data set of SST, MLD and PEX ($1 \times 1^\circ$ grid). SST data were obtained from the World Ocean Atlas 2001 (Conkright et al., 2002), the MLD was calculated as described above. PEX was calculated from satellite-derived primary production data obtained from the Goddard Earth Sciences Data and Information Services Center Distributed Active Archive Center. We used Moderate Resolution Imaging Spectroradiometer (MODIS) data available as 8-day composites (<http://daac.gsfc.nasa.gov/MODIS/Terra/ocean/MOAPW1.shtml>) and calculated monthly means for the time interval December 2002 to November 2003. To estimate export production in 1000 m water depth (PEX) the following equation was applied (Antia et al., 2001):

$$F_{\text{Corg}} = 0.1 \text{PP}^{1.77} z^{-0.68} \quad (2)$$

where F_{Corg} is the organic carbon flux ($\text{g C m}^{-2} \text{yr}^{-1}$) at depth z (m) and PP is primary production ($\text{g C m}^{-2} \text{yr}^{-1}$). Using Eq. (2) should be considered as a first approximation, since the equation is based on annually averaged fluxes, but

export ratios vary on shorter timescales due to seasonally changing productivity patterns (Antia, personal communication).

Calculated monthly fluxes for the 18 foraminiferal species considered (see Fig. 1b) were annually averaged and also converted into relative abundances to allow for a comparison of the model results with core-top foraminiferal fauna data. This implies that months with missing flux calculations due to missing PEX data do not yield any foraminiferal fluxes. Species fluxes were summed to assess total foraminiferal fluxes (note that “total” here means only the sum of the species included in the model). For the global model output we determined species richness as the number of species present. In addition, species diversity (H), which takes into account the relative abundance p_i of each species, was also determined after Shannon and Weaver (1949):

$$H = - \sum p_i \ln(p_i). \quad (3)$$

H increases with increasing species richness and with more evenly distributed relative abundances of the species. For a better comparison between the ocean basins, we combined the pink and white varieties of *G. ruber* when calculating species richness and diversity, since *G. ruber* (pink) occurs only in the Atlantic Ocean.

Maps illustrating our results were generated using the software Ocean Data View (Schlitzer, 2002a).

Table 3. Adjusted coefficients of determination (r^2) and standardized regression coefficients of the multiple linear regressions between the transformed variables SST*, MLD*, PEX* and FLX* for the 18 species included in the model.

Species	Adj. r^2	SST* β_1	MLD* β_2	PEX* β_3
<i>G. bulloides</i>	0.500	0.419	0.175	0.452
<i>G. calida</i>	0.400	0.494	0.243	0.196
<i>G. siphonifera</i>	0.538	0.723	0.162	0.091
<i>G. glutinata</i>	0.352	0.316	0.208	0.397
<i>G. ruber</i> (pink)	0.537	0.547	0.372	0.226
<i>G. ruber</i> (white)	0.696	0.836	0.117	0.188
<i>G. sacculifer</i>	0.541	0.676	0.154	0.195
<i>G. inflata</i>	0.443	0.534	0.307	0.239
<i>G. menardii</i>	0.419	0.563	0.210	0.309
<i>G. scitula</i>	0.425	0.522	0.269	0.189
<i>G. truncatulinoides</i>	0.337	0.338	0.394	0.083
<i>G. rubescens</i>	0.290	0.463	0.289	0.077
<i>N. dutertrei</i>	0.445	0.457	0.191	0.399
<i>N. pachyderma</i> (dex.)	0.750	0.772	0.144	0.253
<i>N. pachyderma</i> (sin.)	0.728	0.946	0.233	0.071
<i>O. universa</i>	0.326	0.439	0.197	0.174
<i>P. obliquiloculata</i>	0.298	0.519	0.252	0.200
<i>T. quinqueloba</i>	0.685	0.808	0.193	0.099

Highest β -value for each species is marked bold

2.4 Comparison to coretop data

To compare our model results with an independent data set we used the Brown University Foraminiferal Database (Prell et al., 1999) (ftp://ftp.ngdc.noaa.gov/paleo/paleocean/brown_foram/). This data set contains foraminifera counts of 1265 globally distributed coretop samples. We calculated relative abundances of the species, species richness and the Shannon diversity index by considering only the 18 species included in our model.

2.5 Comparison to sediment trap data not included in the calibration

Several sediment trap data have not been included in the calibration data set because they were lacking the organic carbon flux data and hence were incomplete. These data (for details and references see Table 2) were used to compare measured with modeled foraminiferal fluxes. To account at least in part for interannual differences in environmental parameters, the model was forced with monthly SST data covering the time of sediment-trap deployment that were obtained from the IGOSS database (Reynolds and Smith, 1994) instead of using long-term monthly means from the World Ocean Atlas 2001 (Conkright et al., 2002).

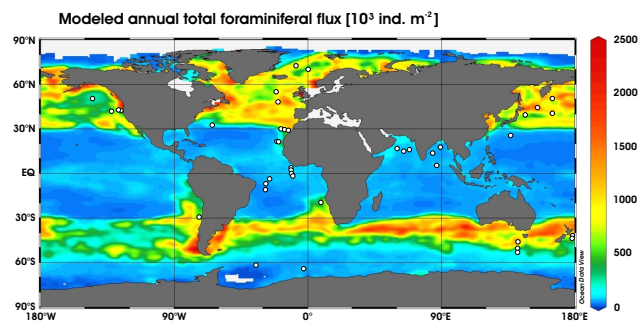


Fig. 2. Modeled annual total foraminiferal flux [10^3 ind. m^{-2}] of the 18 species included in the model, circles mark positions of sediment traps comprised in the calibration data set (see Table 1).

3 Results

3.1 Statistical analysis

Table 3 shows the results of the multiple linear regressions of the transformed variables conducted for every species. Adjusted r^2 values lie between 0.29 for *G. rubescens* and 0.75 for *N. pachyderma* (dex.), indicating that variations in SST, MLD and PEX explain between ~ 30 to 75% of the recorded foraminiferal fluxes. The standardized regression coefficients reflect the significant influence of SST on most of the species, especially on *G. siphonifera*, *G. ruber* (white), *G. sacculifer*, *N. pachyderma* (dex. and sin.) and *T. quinqueloba*. Export production seems to play an important role primarily for *G. bulloides*, *G. glutinata*, *G. menardii* and *N. dutertrei*, while regression coefficients for the MLD are highest for *G. ruber* (pink), *G. inflata* and *G. truncatulinoides*.

3.2 Annual species abundances – global model experiment vs. coretops

The modeled annual total foraminiferal flux is shown in Fig. 2. Low fluxes were calculated across the tropics and in polar regions. A wide band of high foraminiferal fluxes can be seen in the northern hemisphere ($>30^\circ$ N), a comparably narrow band of highest fluxes in the Southern Ocean (predominantly between 30 – 45° S). Increased planktic foraminiferal fluxes were also predicted within the major Eastern Boundary Currents.

Figure 3 illustrates species richness calculated from the global model output as compared to the richness monitored in surface sediments. The general trend of highest species richness at mid-latitudes and decreasing values towards higher latitudes predicted by the model is also reflected in foraminiferal assemblages of coretops. In addition the model reproduces higher species richness in low latitudes of the eastern Atlantic compared to the western part as observed in surface sediments. Furthermore, the distribution pattern of species diversities H calculated for the model

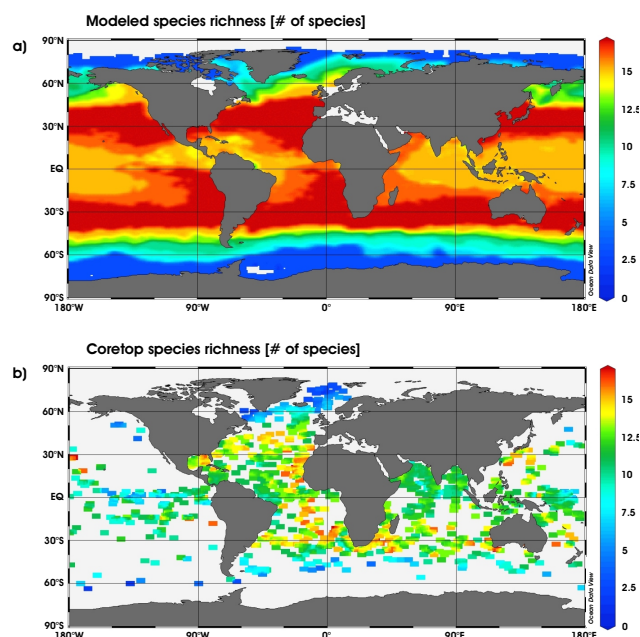


Fig. 3. Species richness [# of species] calculated from (a) the model and (b) coretop foraminiferal assemblages (Prell et al., 1999) considering only species included in the model. White and pink *G. ruber* are combined.

output after Shannon and Weaver (1949) resembles coretop diversity patterns (Fig. 4). In general, species diversities are distinctly lower in high latitudes and show maximum values in rather narrow bands at mid-latitudes in surface sediments and model results. A shift of high diversities to lower latitudes detected in eastern Atlantic coretop samples is predicted by the model as well.

The comparison of modeled annual relative abundances of individual species with their abundances in surface sediments revealed that the global abundance patterns of some species match favorably well with their seafloor record while others are not properly reproduced. For example, Figs. 5–8 show modeled and coretop relative abundances of the four species *N. pachyderma* (sin.), *N. pachyderma* (dex.), *G. bulloides* and *G. siphonifera*, respectively, which were among the species better predicted in the model. Coretop as well as modeled assemblages yield highest abundances of the cold-water species *N. pachyderma* (sin.) in polar waters. Modeled *N. pachyderma* (dex.) shows increased contribution to the foraminiferal assemblage around 40° N/S, following the Gulf Stream into the northeastern N-Atlantic and being present in significant amounts in the major upwelling areas along the Eastern Boundary Currents, just as recorded by surface sediments. Peak abundances of *G. bulloides* are modeled at mid-latitudes which is mirrored by coretops as well. However, relative abundances are being significantly overestimated in the N-Atlantic and underestimated in highly productive upwelling areas like the western Arabian Sea or off

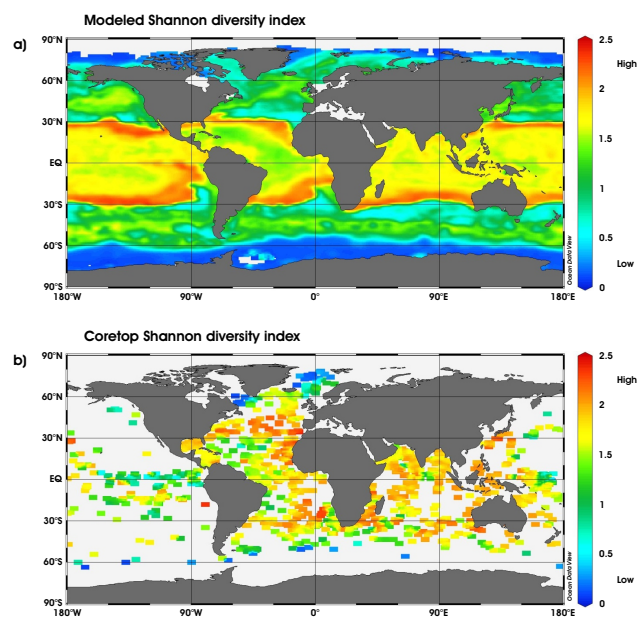


Fig. 4. Species diversity H after Shannon and Weaver (1949) calculated from (a) the model and (b) coretop foraminiferal assemblages (Prell et al., 1999) considering only species included in the model. White and pink *G. ruber* are combined. Higher values correspond to higher species diversity.

NW-Africa. In turn, calculations for *G. siphonifera* show maximum abundances in low latitudes of the oligotrophic subtropical gyres especially in the W-Pacific, while Eastern Boundary regions and the eastern equatorial Pacific yield reduced abundances of this species, thus being in good agreement with the seafloor record. In contrast, Figs. 9 and 10 illustrate two species (*N. dutertrei* and *O. universa*) whose modeled global distribution patterns are not consistent with those of surface sediments. Even though the contribution of *N. dutertrei* to the planktic foraminiferal assemblage is limited to predominantly low latitudes in both model and coretops, modeled peak abundances are found along continental margins, which is not supported by surface sediment data. In turn, maximum coretop abundances in the eastern equatorial Pacific are not reproduced by the model. *O. universa* shows highest modeled abundances in major upwelling areas, which is in contrast to coretop data. Global distributions of modeled and coretop relative abundances of the other 12 species included in the model can be found in Appendix B (Figs. B1–B12).

3.3 Seasonal variations of foraminiferal flux – model vs. sediment traps

To test the predictive skills of the model on a seasonal basis, modeled and measured foraminiferal fluxes were compared for sediment traps not included in the calibration data set. Figures 11 and 12 demonstrate exemplarily results on

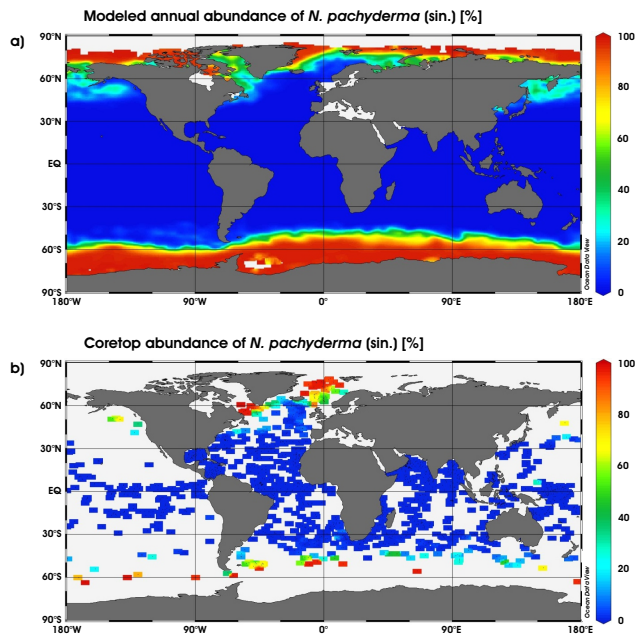


Fig. 5. (a) Modeled annual abundance of *N. pachyderma* (sin.) [%], (b) coretop abundance of *N. pachyderma* (sin.) [%] (Prell et al., 1999). Only species included in the model were considered.

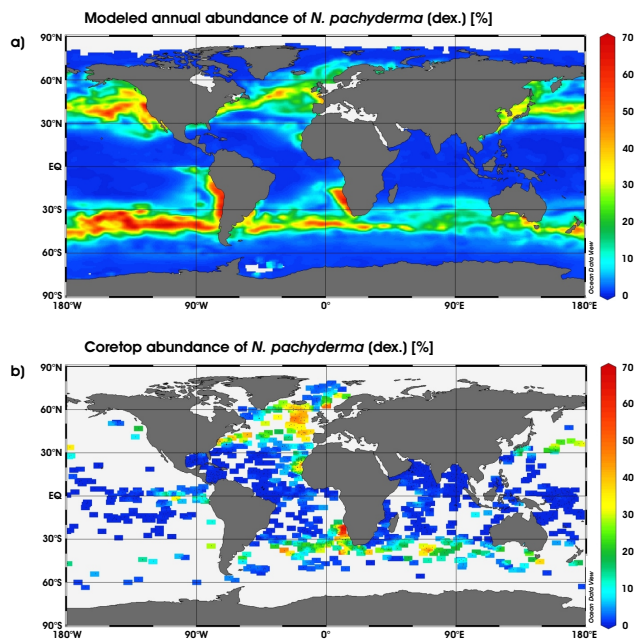


Fig. 6. As in Fig. 5 but for *N. pachyderma* (dex.).

some species for two sediment traps from the Cariaco Basin and the Somalia upwelling, respectively (Conan and Brummer, 2000; Tedesco and Thunell, 2003; see Table 3). One of the principal results visible in the figures is that absolute foraminiferal fluxes are not properly reproduced by the model. In most cases modeled species fluxes are signifi-

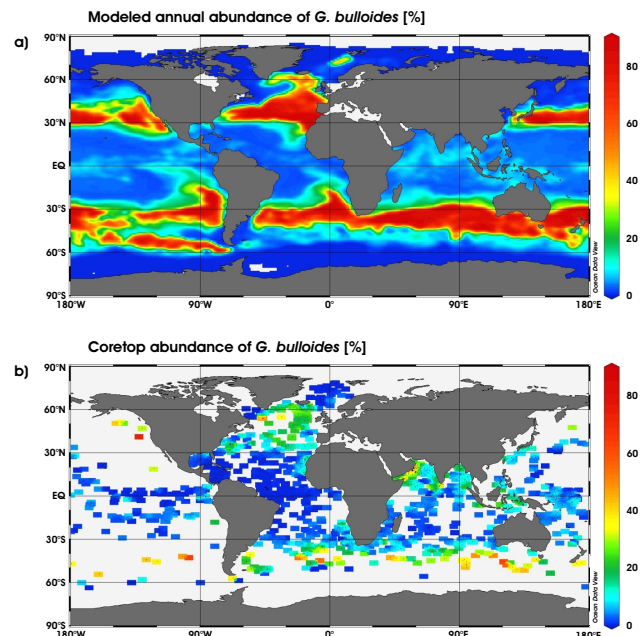


Fig. 7. As in Fig. 5 but for *G. bulloides*.

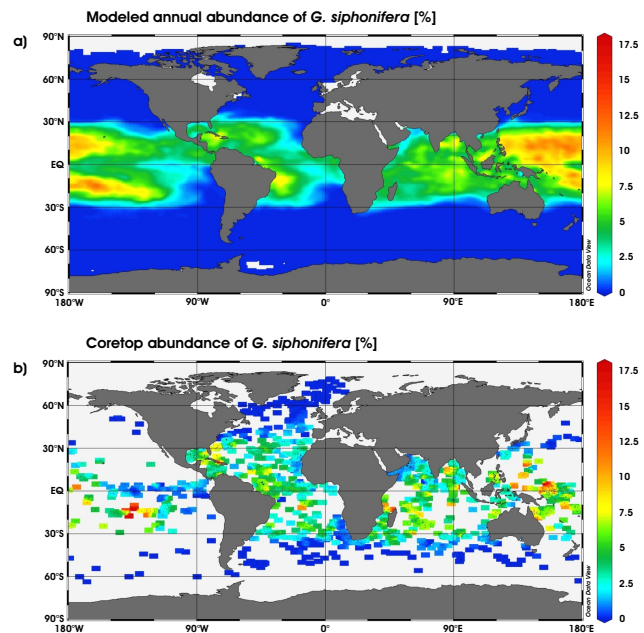


Fig. 8. As in Fig. 5 but for *G. siphonifera*.

cantly underestimated compared to trap-derived fluxes. Regarding the seasonal pattern of species fluxes we observe some species where predicted fluxes match fairly well with their trap record (e.g. *O. universa* and *G. ruber* (white) in Fig. 11, *T. quinqueloba* and *G. glutinata* in Fig. 12), while other species at the same stations seem to be out of phase (e.g. *G. menardii* in Fig. 11 and *G. sacculifer* in Fig. 12). Interannual variability in the exact timing or magnitude of peak

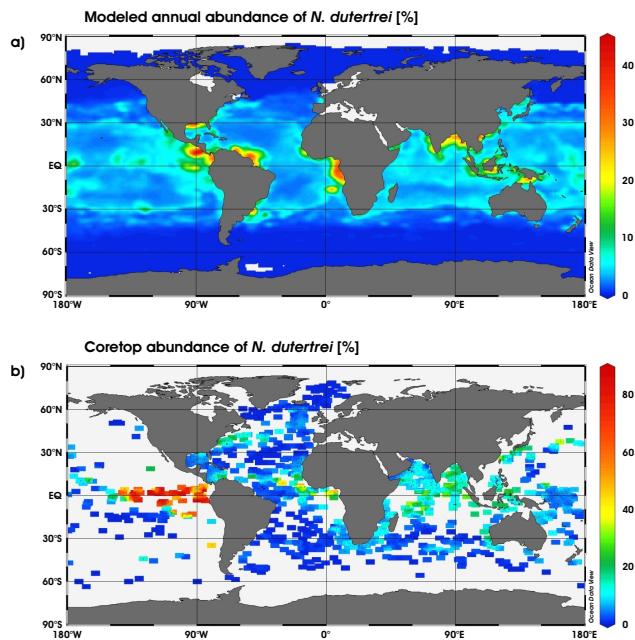


Fig. 9. As in Fig. 5 but for *N. dutertrei*. Note different scaling of color bars.

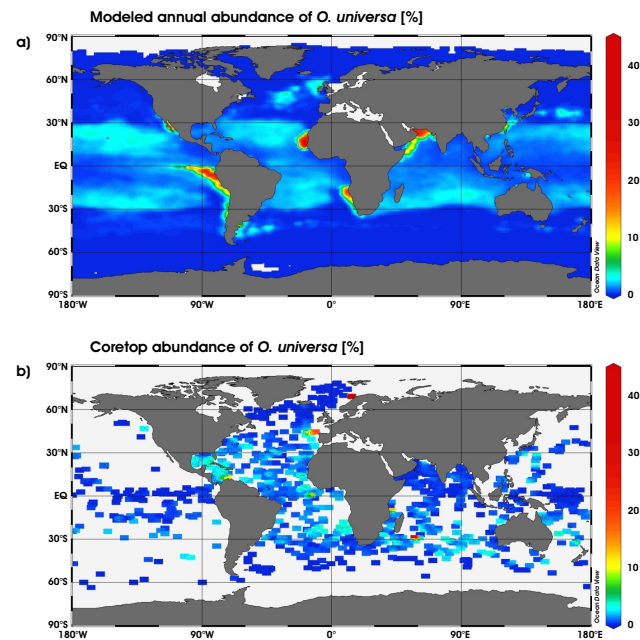


Fig. 10. As in Fig. 5 but for *O. universa*.

fluxes is not reproduced by the model, which can clearly be seen in *T. quinqueloba* and *O. universa* fluxes in the Cariaco Basin (Fig. 11). It should be noted, however, that such variability is not properly taken into account in the model due to the forcing, which is partly based on climatological data (MLD) or a single year (PEX; see discussion).

4 Discussion

4.1 Model output vs. observations from coretops and sediment traps

The empirical model presented here is, to our knowledge, the first attempt to predict planktic foraminiferal fluxes on a global scale. Many principal distribution patterns of foraminiferal species recognized from the model experiments do match observations in coretops or sediment traps, even though the model still produces problematic outputs in many places.

4.1.1 Total foraminiferal flux, species richness and diversity

Annually averaged total foraminiferal fluxes calculated by the model are highest in temperate and subpolar waters and low in polar waters and the tropics with minima in the centers of the oligotrophic subtropical gyres reflecting planktic foraminiferal needs for sufficient food supply (Fig. 2). Higher total foraminiferal fluxes were also modeled in lower latitudes associated with Eastern Boundary Currents charac-

terized by strong coastal upwelling and hence higher productivity. An increase in fluxes, though rather small, was also calculated for the seasonal upwelling region in the western Arabian Sea and in the equatorial upwelling regions of the Atlantic and Pacific Oceans.

Modeled meridional gradients in species richness and diversity compare favorably with coretop data (Figs. 3 and 4) and are consistent with the general trend of an overall decrease in faunal diversity with increasing latitude (e.g. MacArthur, 1965; Stehli et al., 1969). Analyzing planktic foraminiferal diversity from surface water samples Ottens and Nederbragt (1992) showed that deviations from this global trend are related to specific ocean environments as suggested earlier by Stehli (1965). They demonstrated that water mass boundaries or frontal zones are characterized by high diversity of planktic foraminiferal faunas due to mixing of species from adjacent water masses. On the other hand, they found that variable environments with distinct seasonal or short-term variations in oceanic parameters like in upwelling areas host relatively low diversity faunas as compared to the surrounding environment, which is in good agreement with our model results. Lower absolute values in coretop species richness particularly in the Pacific Ocean might in part be due to selective dissolution of the surface sediments (Berger, 1968; Thunell and Honjo, 1981; Boltovskoy, 1994; Le and Thunell, 1996; Dittert and Henrich, 2000; Conan et al., 2002), as they are exposed to the surrounding seawater much longer than the trap material, which was used in the model calibration.

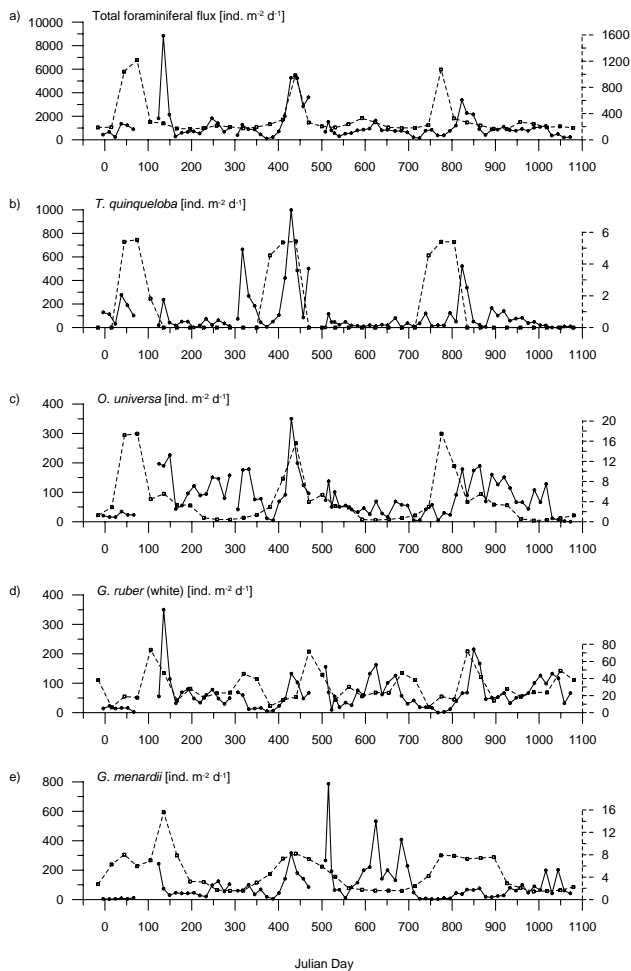


Fig. 11. Comparison of modeled and measured foraminiferal fluxes in $[\text{ind. m}^{-2} \text{ d}^{-1}]$. Solid lines/y-axes and filled circles show fluxes measured in the sediment trap in the Cariaco Basin (Tedesco and Thunell, 2003). Dashed lines/y-axes and open squares show fluxes calculated by the model. Note that in this case the model was forced with actual monthly sea-surface temperatures from the time of trap deployment to account at least in part for interannual differences in environmental parameters (see Sect. 2.5). Trap data are plotted 2 weeks prior to their catchment intervals to account for the time-lag applied in the calibration. (a) Total foraminiferal flux considering only species included in the model, (b) *T. quinqueloba*, (c) *O. universa*, (d) *G. ruber* (white), (e) *G. menardii*, Julian Day 1 = 1 January 1997.

4.1.2 Relative species abundances

Analyzing annual relative abundances of foraminiferal species has shown that particular patterns in the distribution of some species on the seafloor can also be reproduced by the model. Among these is for example the restriction of significant relative abundances of the cold-water species *N. pachyderma* (sin.) to higher latitudes, where it can account for more than 90% of the assemblage (Fig. 5). For

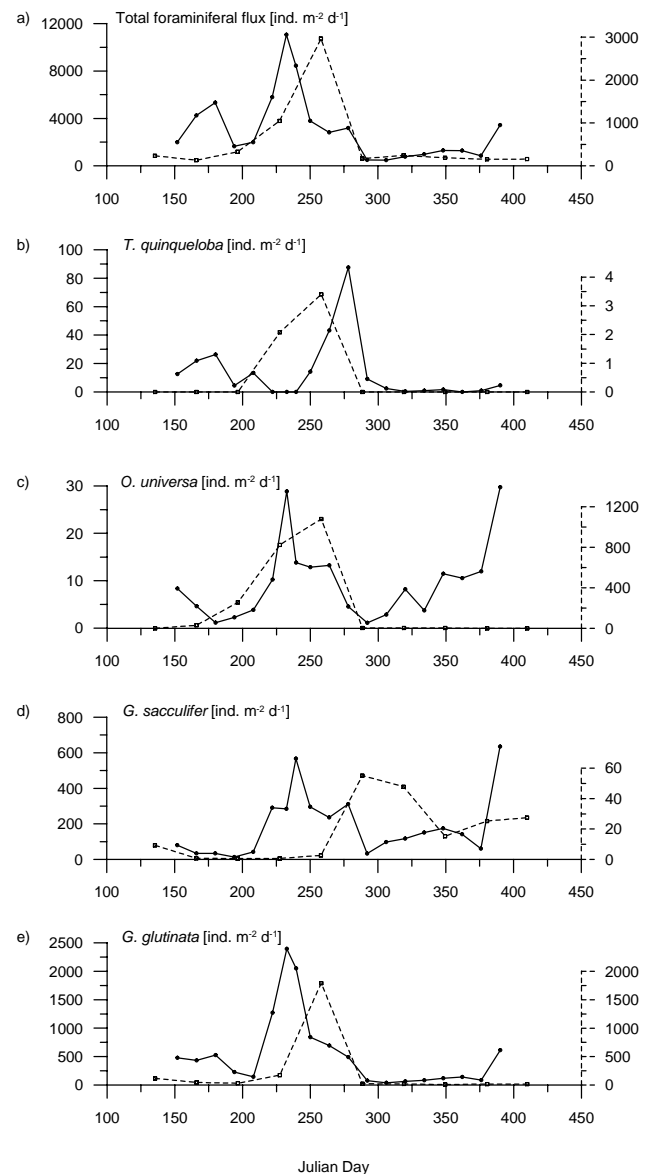


Fig. 12. Comparison of modeled and measured foraminiferal fluxes in $[\text{ind. m}^{-2} \text{ d}^{-1}]$. Solid lines/y-axes and filled circles show fluxes measured in the sediment trap MST8-B in the Somalia Upwelling (Conan and Brummer, 2000). Dashed lines/y-axes and open squares show fluxes calculated by the model. Trap data are plotted 2 weeks prior to their catchment intervals to account for the time-lag applied in the calibration. (a) Total foraminiferal flux considering only species included in the model, (b) *T. quinqueloba*, (c) *O. universa*, (d) *G. sacculifer*, (e) *G. glutinata*, Julian Day 1 = 1 January 1992.

N. pachyderma (dex.), preferring subpolar to transitional water masses, the modeled relative abundance pattern does not only match the latitudinal distribution but also more complex structures like increased abundances in the major upwelling areas along Eastern Boundary Currents, which are clearly visible in coretops off SW- and NW-Africa (Fig. 6).

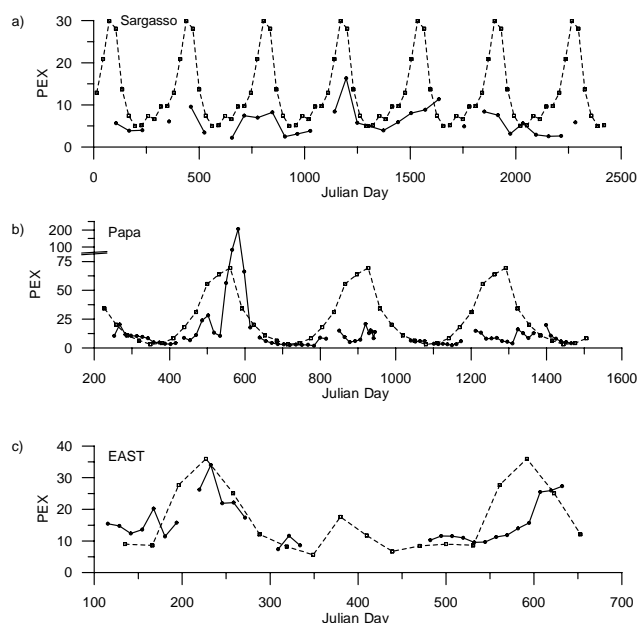


Fig. 13. Comparison of export production values in 1000 m water depth (PEX) calculated for the calibration and the global data set in [$\text{mg C}_{\text{org}} \text{m}^{-2} \text{d}^{-1}$] for the stations (a) in the Sargasso Sea (Julian Day 1 = 1 January 1978), (b) at Ocean Station Papa (Julian Day 1 = 1 January 1982), and (c) in the eastern Arabian Sea (EAST, Julian Day 1 = 1 January 1986). Solid lines and filled circles show PEX calculated from sediment-trap-derived C_{org} fluxes as applied in the model calibration (for references see Table 1). Dashed lines and open squares show PEX calculated from satellite-derived primary productivity as applied for the global model run after Antia et al. (2001). Trap data are plotted 3 weeks prior to their catchment intervals to account for the time-lag applied in the calibration.

Even the higher abundances spreading westward along the eastern equatorial Pacific could be reproduced by the model. As observed from surface sediments, this region and major coastal upwelling areas yield reduced relative abundances of the warm-water species *G. siphonifera*, which shows high abundances in the oligotrophic subtropical gyres (highest in the W-Pacific). This distribution pattern also compares favorably well with our model results (Fig. 8) as does the latitudinal distribution of *G. bulloides* abundances (Fig. 7). In contrast, increased relative abundances of *G. bulloides* in some major upwelling areas (e.g. the western Arabian Sea off Somalia) are not mirrored in the model in spite of a high standardized regression coefficient for PEX^* for this species (Table 3). A possible reason might be the bad coverage of very high export productivities in the calibration data set. Only 21 of the 1327 samples yield PEX values above $40 \text{ mg C}_{\text{org}} \text{m}^{-2} \text{d}^{-1}$ in 1000 m water depth, which is apparently insufficient to reliably predict foraminiferal fluxes in highly productive upwelling regions. Other examples, in which modeled species distributions significantly deviate from their seafloor record are abundance patterns of *N. dutertrei* and *O. uni-*

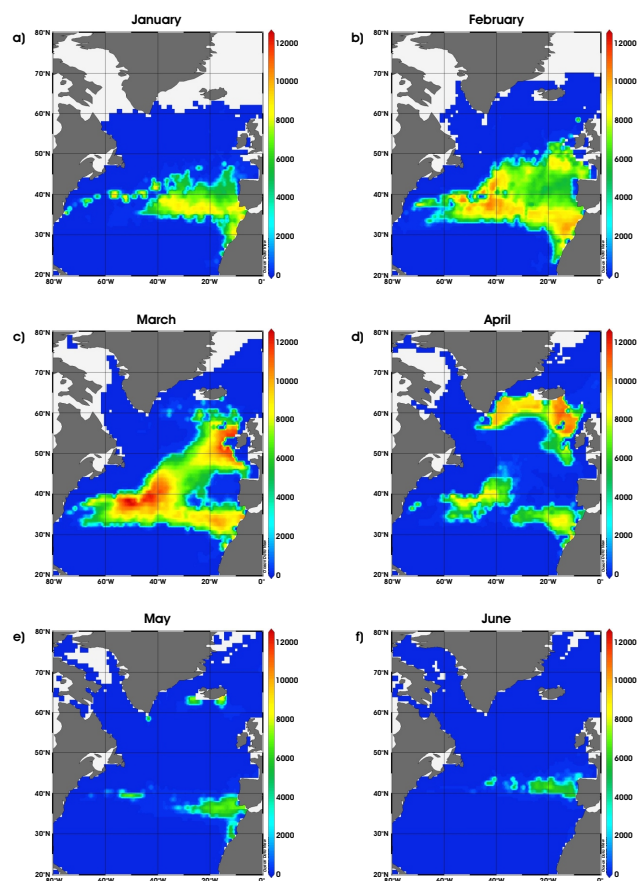


Fig. 14. Modeled monthly fluxes [$\text{ind. m}^{-2} \text{d}^{-1}$] of the species *G. bulloides* in the N-Atlantic for the months January to June (a–f).

versa (Figs. 9 and 10). Even though both model and coretops show minimal abundances of *N. dutertrei* in higher latitudes, higher abundances from the model do not match observations from coretops and vice versa. The apparent mismatch in the eastern equatorial Pacific might in part be due to a lack of calibration samples from that very special ocean environment.

However, when comparing modeled relative abundances of foraminiferal species with coretop distributions it has to be borne in mind, that the model calculates absolute species fluxes from the environmental input parameters. Hence, the quality of the estimate of relative abundances depends on the performance of *all* species in the model, so that fluxes of a poorly represented species worsen the results of better reproduced species as well. Surface sediments, on the other hand, can reflect foraminiferal assemblage variations averaged over several decades or several hundred years (depending on sedimentation rates). Such long-term variations are not represented in the model due to the forcing being used. In addition, sedimentary assemblages may be altered by selective dissolution, by displacement through bottom currents or by bioturbation processes (Boltovskoy, 1994). Furthermore, coretops reveal only relative abundances of foraminiferal

species and do not allow conclusions on absolute species fluxes.

4.1.3 Absolute species fluxes

The lack of absolute shell fluxes of foraminiferal species on a global scale makes it difficult to assess the corresponding model data. Bé and co-workers analyzed the geographic distribution of various species in the Atlantic and Indian Oceans using plankton tows, and presented absolute and relative abundances of the planktic foraminiferal standing stocks (Bé and Hamlin, 1967; Bé and Tolderlund, 1971; Bé and Hutson, 1977). But several reasons argue against a comparison of our model results to their distributional maps. First of all, they used plankton nets with a mesh size of 200 μm in contrast to the 125 and 150 μm sieve size used in the sediment trap studies. Moreover, plankton tow samples represent only snapshots of the foraminiferal assemblages (Boltovskoy et al., 1996) and finally, values of foraminiferal standing stocks cannot be directly translated into downward fluxes of planktic foraminiferal species.

The only alternative to independently test the predictive skills of the model regarding absolute fluxes of planktic foraminifera were several sediment trap studies that were not included in the calibration data set (Table 2, Figs. 11 and 12). One of the principal model results in this context is the significant underestimation of absolute foraminiferal abundances in most cases. This might in part be due to the fact that actual foraminiferal fluxes and relative abundances at certain environmental conditions can be highly variant, as was shown for surface sediment as well as sediment trap databases (Hilbrecht, 1996; Žarić et al., 2005). This variability is not accounted for by the model, which calculates only a single “average” flux for a given set of environmental parameters. These average values are usually comparably small especially because the data set contains a high number of “void observations” (see methods Sect. 2.1). Despite the fact that absolute foraminiferal fluxes are not adequately predicted by the model in most cases, the seasonal signal of species fluxes compares fairly well to sediment trap records for some species. For example, high fluxes of *T. quinqueloba*, *O. universa* and *G. ruber* (white) in the Cariaco Basin (Fig. 11) and of *G. glutinata* in the Arabian Sea (Fig. 12) occur at similar times in the model as they were in fact recorded by the sediment traps. At the same time other species like *G. menardii* in the Cariaco Basin and *G. sacculifer* in the Somalia upwelling are sometimes out of phase in the model. Further species have barely any seasonality in fluxes even though the sediment trap record shows high peak fluxes at certain times (e.g. *G. bulloides* in the Cariaco Basin, not shown). Considering all species on a global scale, we could not detect any global systematic error in the predicted fluxes neither regarding their magnitude nor their seasonality for any species.

4.2 Pitfalls and potential of the model

Some of the discrepancies between modeled and observed foraminiferal flux patterns may be due to an insufficient calibration of the model. For an optimal calibration it would be necessary to have actual hydrographical as well as productivity data on timescales at which the sediment traps operate (in the order of weeks). Such data were only available for SST (Reynolds and Smith, 1994). The MLD had to be calculated from climatological temperature and salinity data (Conkright et al., 2002). Thus, information on interannual differences of the mixed layer is lost, and potential subsequent variations of foraminiferal fluxes are inadequately reflected in the hydrographic parameters of the calibration data set. The same is true when comparing the modeled seasonal signal of foraminiferal assemblages with measured foraminiferal fluxes of sediment traps not included in the calibration. The model was forced with long-term mean values of MLD, and PEX values of an exemplary year, which may deviate significantly from the actual situation that the foraminifera were living in prior to settling into the trap. Again, information on the variability of the environmental input parameters cannot be taken into account, and the model is not able to reproduce interannual changes in foraminiferal fluxes (Fig. 11), which can be significant (e.g. Sautter and Thunell, 1989; Tedesco and Thunell, 2003; Marchant et al., 2004).

Export production estimates introduce the largest uncertainty into the model, because export ratios should vary over the course of the year due to seasonally changing productivity patterns, but so far there are no algorithms on organic carbon transfer efficiencies based on shorter than annual timescales. Moreover, we had to use a different approach for the calculation of PEX in the global data set than for the PEX calculation applied in the calibration (see methods Sects. 2.1 and 2.3). Whereas we could normalize trap-measured organic carbon fluxes to 1000 m in the calibration (Martin et al., 1987; Francois et al., 2002), we had to calculate export production from satellite-derived primary production data of an exemplary year after Antia et al. (2001) in order to obtain monthly PEX values for our global data set. The comparability of both approaches remains an open question. Comparing PEX values from both our calibration as well as our global data set at the positions of the sediment traps shows that both PEX values are comparable at some positions/times but that in many cases satellite-derived values can be significantly higher than trap-derived values for other positions/times (Fig. 13). This means that the model often calculates foraminiferal fluxes at higher export productions than those recorded in the calibration data set for the same position.

As the quantification of downward organic carbon fluxes as well as their relationship to primary production in surface waters are still subject to intensive discussions (e.g. Bueseler, 1998; Laws et al., 2000; Antia et al., 2001; Armstrong et al., 2002; Francois et al., 2002; Lutz et al., 2002;

Schlitzer, 2002b), the export production in 1000 m water depth is probably not well suited as an environmental input parameter for a planktic foraminiferal flux model. We decided to still use export production, because it was the only productivity-related parameter being available in the temporal resolution of sediment trap samples. However, it would be generally useful to replace the PEX variable in the model by a primary-production dependent variable, which would be a more realistic representation of food availability for planktic foraminifera. Low coefficients of determination (r^2 between 0.29 and 0.75) calculated in the statistical model calibration already suggest that the three environmental parameters SST, MLD and PEX used here are not sufficient to describe the fluxes of all planktic foraminiferal species considered and that additional parameters should be included in the model. Numerous studies revealed that productivity often being linked to the nutrient content is a significant factor influencing planktic foraminiferal fluxes and assemblage compositions (e.g. Bé and Hutson, 1977; Ortiz et al., 1995; Watkins et al., 1996; Eguchi et al., 1999; Schiebel et al., 2001; Morey et al., 2005). In this context it would also be reasonable not to correlate species fluxes exclusively to conditions at the sea surface but to consider different water depths as well to include the occurrence of deep chlorophyll maxima that some foraminifera like *N. pachyderma* or *N. dutertrei* have often been associated with at least for part of their life cycles (e.g. Fairbanks and Wiebe, 1980; Fairbanks et al., 1982; Reynolds and Thunell, 1986; Kohfeld et al., 1996).

Other environmental conditions not included in our model so far, that have been supposed to have an impact on planktic foraminiferal fluxes and assemblage compositions are light intensity, circulation patterns or salinity (e.g. Bijma et al., 1990b; Ortiz et al., 1995; Watkins et al., 1996; Guptha et al., 1997). Based on studies of laboratory cultures (Bijma et al., 1990b) and core-top sediments (Morey et al., 2005) that suggest that salinity does not have a significant effect on foraminiferal species distributions in modern oceans, we decided to omit salinity in our model to avoid enhanced multicollinearity, since salinity is often highly correlated with SST.

Several factors potentially contributing to noise or error in the calibration data set have been discussed by Žarić et al. (2005) and shall only briefly be mentioned here. Among these are a patchy distribution of foraminifera in the ocean (e.g. Bé and Hutson, 1977), foraminiferal life cycles (e.g. Bé, 1977; Bijma et al., 1990a, 1994), differences in sinking velocities of foraminiferal shells (e.g. Takahashi and Bé, 1984; Bijma et al., 1994), a lateral component in particle fluxes (e.g. Freudenthal et al., 2001; Wilke et al., 2005¹), the presence of distinct genotypes within certain morphospecies having distinct ecological preferences (e.g. summarized in Kucera and Darling, 2002), the trapping efficiency

of sediment traps (Scholten et al., 2001; Yu et al., 2001) and the taxonomic consistency of the planktic foraminiferal studies used here. Furthermore, in the sediment-trap investigations included here two different sieve sizes were used for foraminiferal analyses (125 and 150 μm , respectively). As was shown by Carstens et al. (1997), species flux and relative abundance can differ significantly when the minimum size of the counted foraminifera is changed. Within the scope of this investigation, however, we have to use every available data set, and most of the data were available in the given size fraction only.

Nevertheless, even though the model presented here produces problematic outputs in some places, many general distribution patterns of planktic foraminifera can be recognized. The potential of such a foraminiferal flux model to be used for paleoreconstructions by identifying seasonal flux signals for certain species shall be illustrated on the following example. Figure 14 shows modeled monthly fluxes (January to June) for the species *G. bulloides* in the N-Atlantic. Peak fluxes are calculated for spring (March) with high fluxes occurring first in more southerly parts around 40° N in January/February and then migrating northward through March and April. Thus our model might confirm results from isotopic studies on NE-Atlantic surface sediments by Ganssen and Kroon (2000), who classified *G. bulloides* as a species typical of the spring bloom rather than reflecting summer temperatures.

The model presented here is static in the sense that it runs independently for every species, grid-point and month. It carries out every calculation based exclusively on “actual” values of environmental parameters, not taking into account previous hydrographic situations or foraminiferal standing stocks. Adding a dynamic component to the model might thus improve it significantly. In that case the new state of the model would be calculated memorizing the current state and adding a certain rate of change according to changes of the environmental parameters. An important step towards this direction would be coupling our model to current ecosystem models as described by Moore et al. (2001, 2004), which could provide necessary state variables of the pelagic ecosystem.

5 Summary and conclusions

(1) The empirical model described here is, to our knowledge, the first attempt to globally predict planktic foraminiferal fluxes at species level depending on the environmental parameters sea-surface temperature, mixed-layer depth and export production. It was calibrated using a combination of sediment trap as well as hydrographic data and forced with a global data set of SST, MLD and PEX to calculate monthly foraminiferal fluxes.

(2) Annually averaged total foraminiferal fluxes calculated by the model peak in temperate and subpolar waters and

¹Wilke, I., Meggers, H., and Bickert, T.: Seasonal distribution and stable oxygen isotope composition of planktic foraminifera off NW-Africa (29° N), Deep-Sea Res. I, submitted, 2005.

are lowest in polar waters and the tropics reflecting planktic foraminiferal needs for sufficient food supply. Many principal distribution patterns of foraminiferal species predicted by the model compare favorably with observations from core-tops, among others meridional gradients in species richness and diversity, increased relative abundances of *N. pachyderma* (dex.) in major upwelling areas, and peak abundances of *G. siphonifera* in oligotrophic subtropical gyres. In contrast, relative abundance patterns of some other species are not properly reproduced by the model.

(3) Comparisons between modeled and observed planktic foraminiferal fluxes revealed that absolute downward fluxes are significantly underestimated in most cases. Nevertheless, modeled seasonal flux variations match fairly well with the sediment trap record for some species. Interannual flux variations could not be properly reproduced, possibly because of a lack of actual environmental data for calibration and forcing purposes. Hence, our study stresses the importance of acquiring actual data on environmental parameters while conducting sediment trap experiments to ensure that the hydrographic situation is properly monitored.

(4) The limited predictive skills of the model suggest that additional environmental information should be considered such as a more realistic representation of food availability at different habitat depths of planktic foraminiferal species. This could be achieved by adding a dynamic component to the model and linking it to an ecosystem model.

Appendix A

Table A1. Sea-surface temperature ranges for the species included in the model. Ranges were derived as outlined in Žarić et al. (2005). Outside of these ranges, modeled foraminiferal species fluxes are set to zero.

Species	SST range	
	from	to
<i>G. bulloides</i>	1.9	31.0
<i>G. calida</i>	9.3	31.0
<i>G. siphonifera</i>	11.9	31.0
<i>G. glutinata</i>	1.9	31.0
<i>G. ruber</i> (pink)	16.4	29.6
<i>G. ruber</i> (white)	9.8	31.0
<i>G. sacculifer</i>	9.7	31.0
<i>G. inflata</i>	1.9	29.6
<i>G. menardii</i>	13.3	30.5
<i>G. scitula</i>	2.0	29.8
<i>G. truncatulinoides</i>	3.5	29.8
<i>G. rubescens</i>	16.3	29.8
<i>N. dutertrei</i>	2.6	31.0
<i>N. pachyderma</i> (dex.)	-1.8	29.8
<i>N. pachyderma</i> (sin.)	-1.8	23.7
<i>O. universa</i>	6.1	29.7
<i>P. obliquiloculata</i>	16.4	30.0
<i>T. quinqueloba</i>	-1.8	25.9

Appendix B

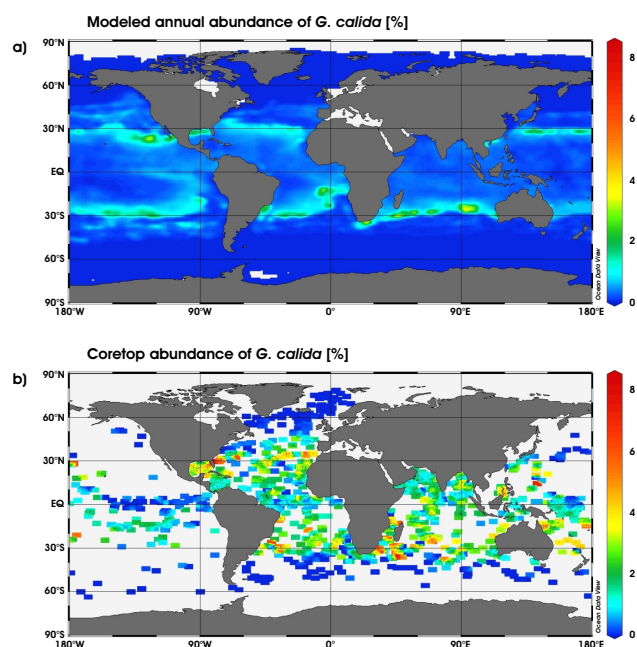


Fig. B1. (a) Modeled annual abundance of *G. calida* [%], (b) coretop abundance of *G. calida* [%] (Prell et al., 1999). Only species included in the model were considered.

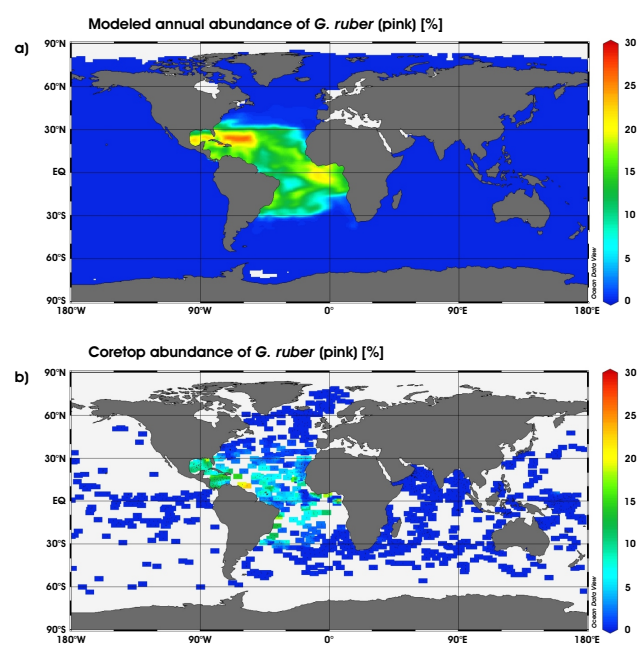


Fig. B3. As in Fig. B1 but for *G. ruber* (pink).

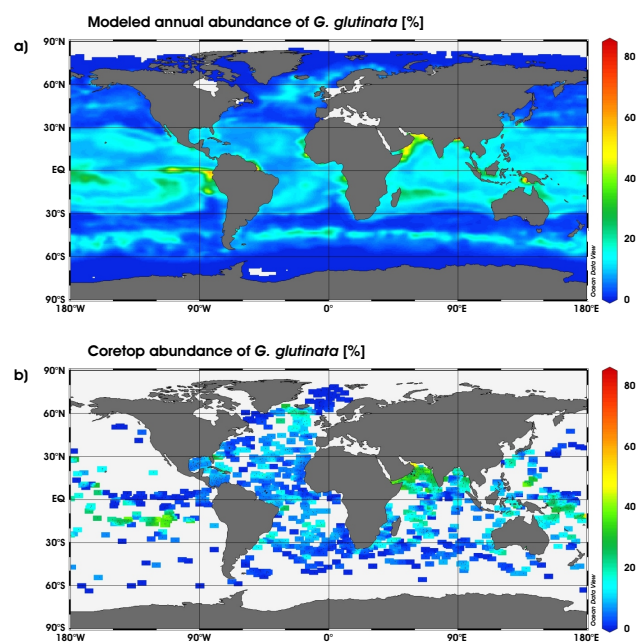


Fig. B2. As in Fig. B1 but for *G. glutinata*.

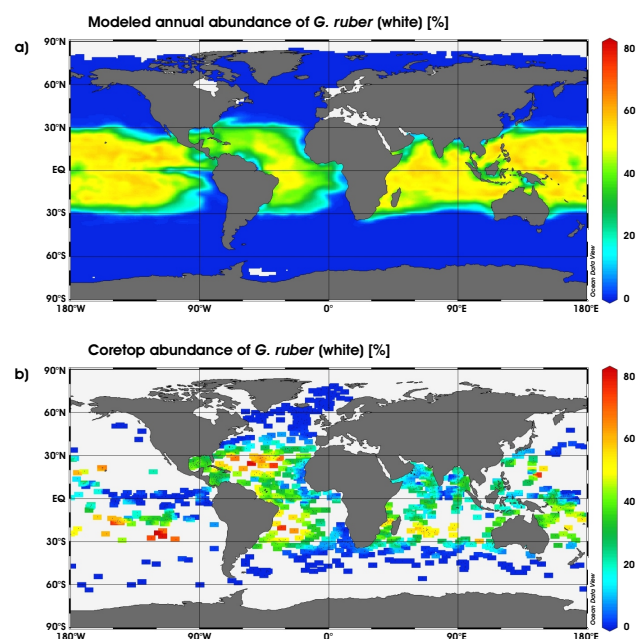


Fig. B4. As in Fig. B1 but for *G. ruber* (white).

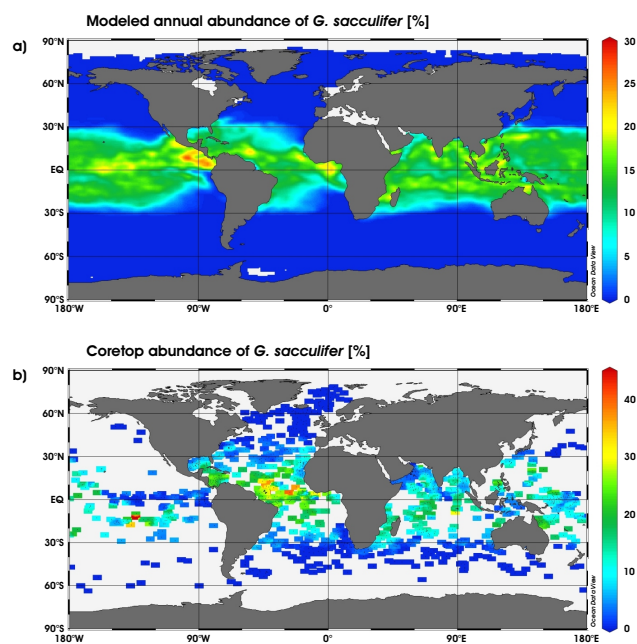


Fig. B5. As in Fig. B1 but for *G. sacculifer*. Note different scaling of color bars.

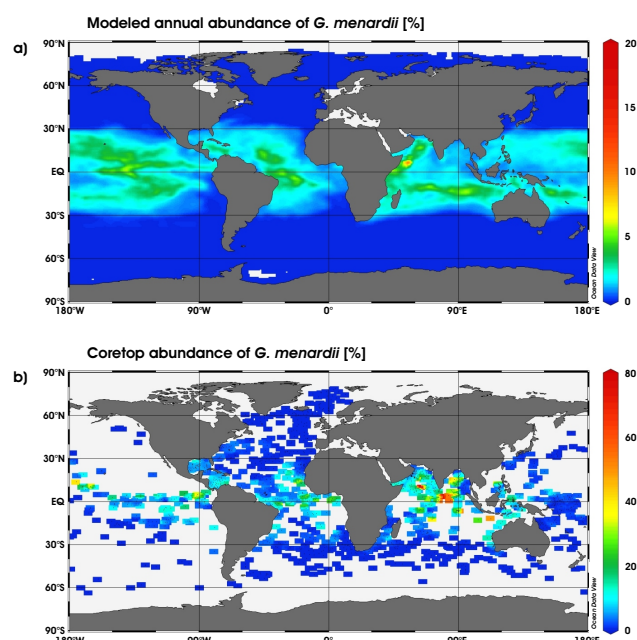


Fig. B7. As in Fig. B1 but for *G. menardii*. Note different scaling of color bars.

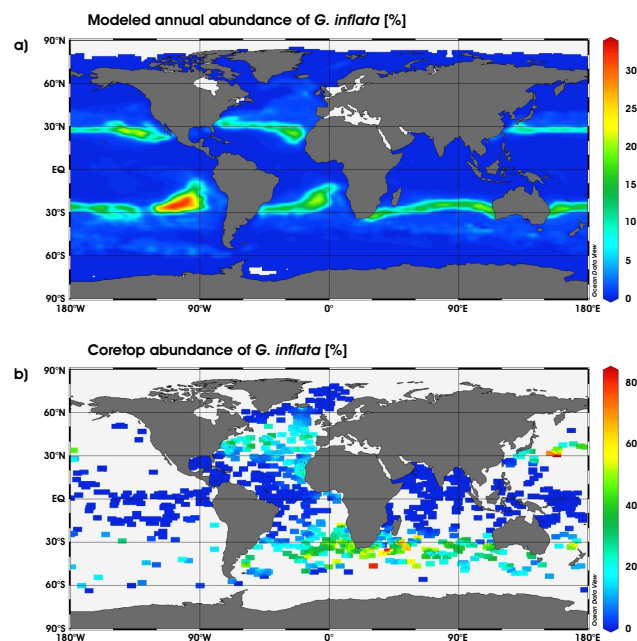


Fig. B6. As in Fig. B1 but for *G. inflata*. Note different scaling of color bars.

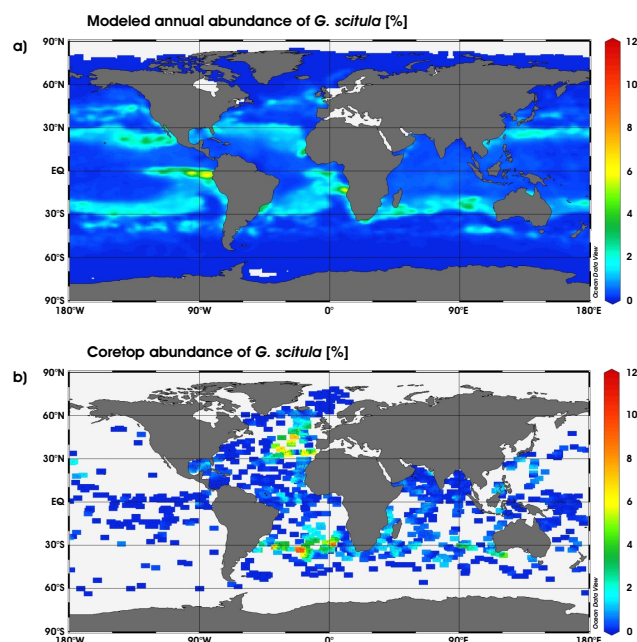


Fig. B8. As in Fig. B1 but for *G. scitula*.

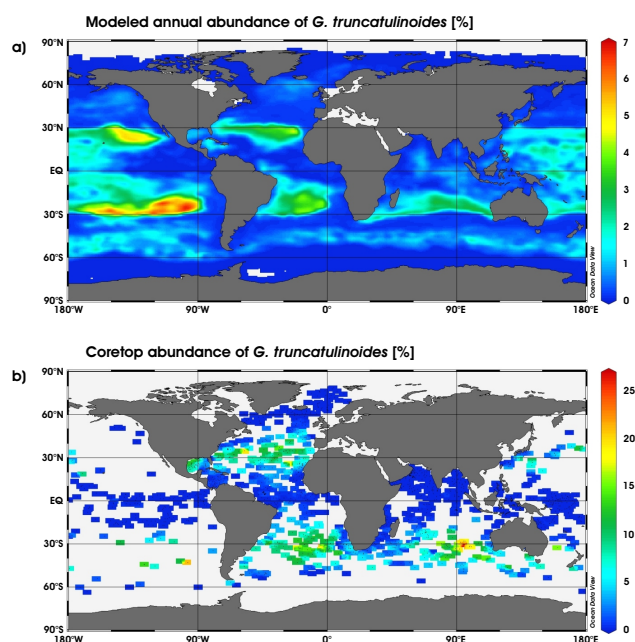


Fig. B9. As in Fig. B1 but for *G. truncatulinoides*. Note different scaling of color bars.

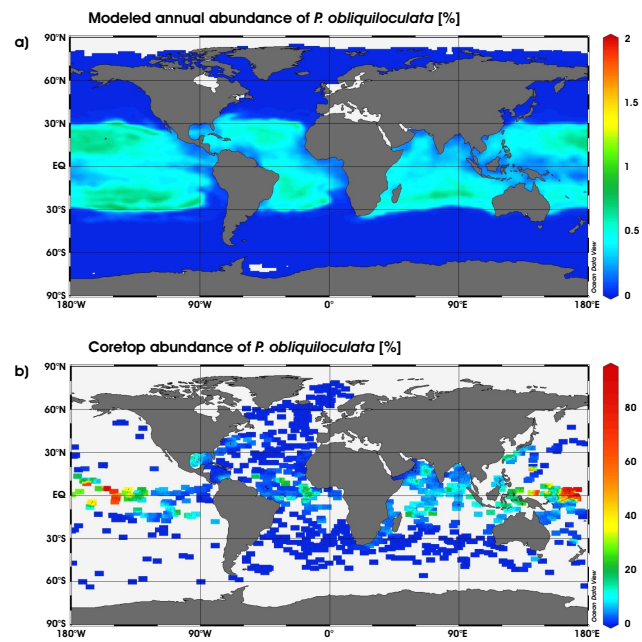


Fig. B11. As in Fig. B1 but for *P. obliquiloculata*. Note different scaling of color bars.

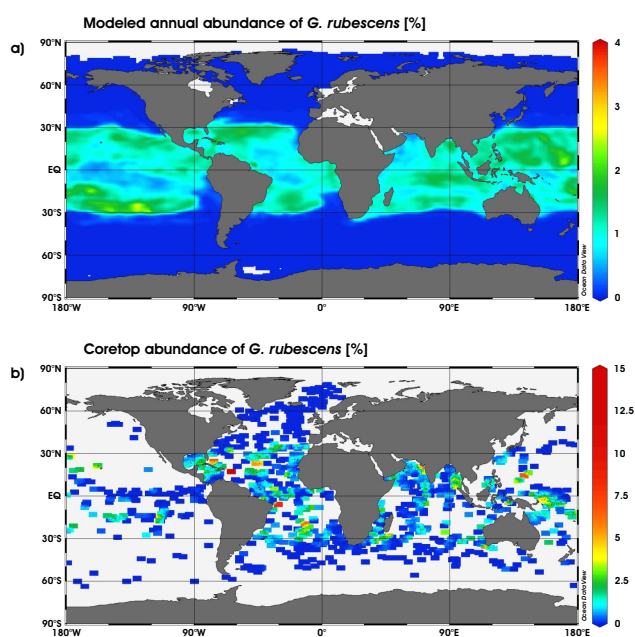


Fig. B10. As in Fig. B1 but for *G. rubescens*. Note different scaling of color bars.

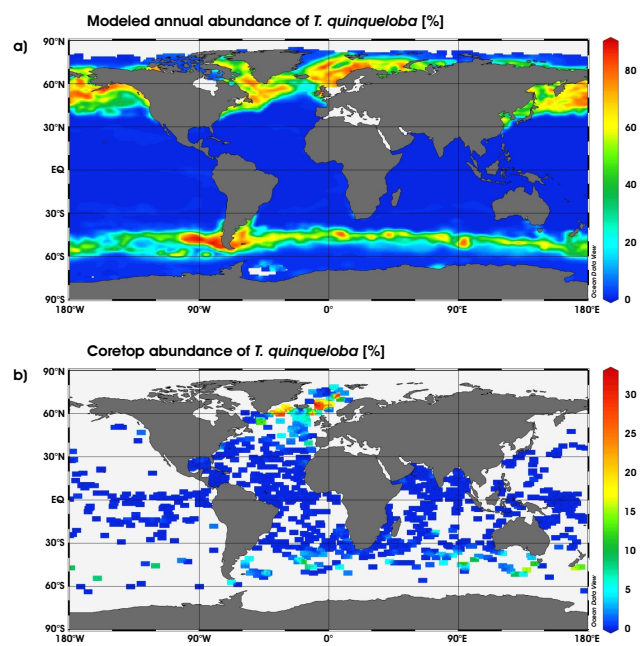


Fig. B12. As in Fig. B1 but for *T. quinqueloba*. Note different scaling of color bars.

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